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Review article

From the exposome to mechanistic understanding of chemical-induced adverse effects

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ABSTRACT

The exposome encompasses an individual's exposure to exogenous chemicals, as well as endogenous chemicals that are produced or altered in response to external stressors. While the exposome concept has been established for human health, its principles can be extended to include broader ecological issues. The assessment of exposure is tightly interlinked with hazard assessment. Here, we explore if mechanistic understanding of the causal links between exposure and adverse effects on human health and the environment can be improved by integrating the exposome approach with the adverse outcome pathway (AOP) concept that structures and organizes the sequence of biological events from an initial molecular interaction of a chemical with a biological target to an adverse outcome. Complementing exposome research with the AOP concept may facilitate a mechanistic understanding of stress-induced adverse effects, examine the relative contributions from various components of the exposome, determine the primary risk drivers in complex mixtures, and promote an integrative assessment of chemical risks for both human and environmental health.

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Abbreviations: AEP, Aggregate exposure pathway; AO, Adverse outcome; AOP, adverse outcome pathway; EWAS, exposome-wide association studies; KE, key event; HTS, high throughput screening; MIE, molecular initiating event; TD, toxicodynamic; TK, toxicokinetic.

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1. Introduction

The exposome expands our perception of lifetime exposure because it integrates exogenous chemicals with genetic and external factors that generate chemicals inside the body and thereby may pose threats to human health (Miller and Jones, 2014; Rappaport and Smith, 2010; Wild, 2012). The external contribution to the human exposome is determined by environmental exposure, also termed the eco-exposome (Lioy and Smith, 2013), such as exposure via air, food, water, dust, and use of consumer products (Fig. 1). Apart from environmental pollutants and their biotransformation products, the exposome includes endogenous metabolites and markers of the adaptive cellular stress responses, as well as chemicals that are generated in response to psychosocial stress and lifestyle factors. These joint exposures can be related to adverse health effects via exposome-wide association studies (EWAS;

Rappaport, 2012) without attempting to identify mechanistic causes (Fig. 1). Importantly, these associations capture the joint effect of many stressors acting in concert, which invokes mixture effects not only in chemical space of exogenous and endogenous compounds, but also mixtures in time, including the time dependence of effects. The exposome has thus been advocated as a key to cumulative risk assessment (Smith et al., 2015).

During the last decade, the exposome approach has mainly been considered in epidemiology, while the complementary concept of Adverse Outcome Pathways (AOP) has emerged in (eco)toxicology (Ankley et al., 2010). The AOP concept links the exposure of chemicals to their cellular concentrations and molecular initiating events (MIE), through network/pathway disturbances and key events (KE) to responses at the cellular, organ, organism and, finally, population and ecosystem levels (Fig. 1). The AOP concept aims to enhance the utility of

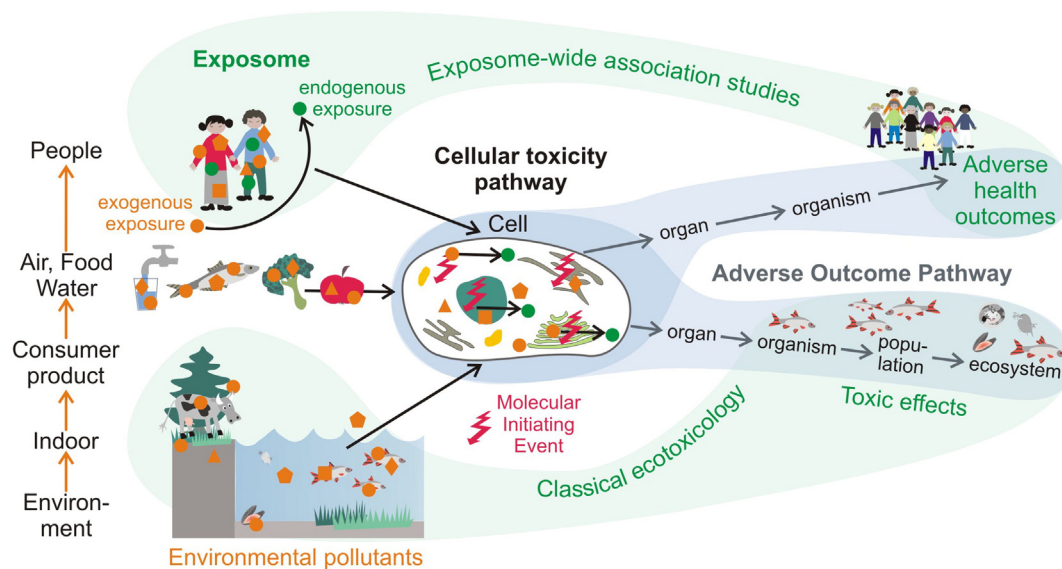


Fig. 1. Multiple chemical exposures of the environment and their link via environmental media and the food chain to human exposure. Any type of exogenous chemical exposure will change the endogenous exposure, both of which will elicit effects on cellular toxicity pathways. The cellular level might serve as integrator to understand both, the pathways to adverse health outcomes as well as to ecosystem-level effects.

mechanistic data for understanding and predicting adverse effects. It also aligns in this goal with systems toxicology (Sturla et al., 2014) and the Tox21 program, a joint initiative of the US National Institute of Environmental Health Sciences (NIEHS) and the US Environmental Protection Agency (EPA) (Betts, 2013; National Research Council, 2007). While the concepts of AOP and cellular toxicity pathways (Patlewicz et al., 2013) account for multiple pathways triggered by a chemical or stressor and are per definition chemical agnostic, most examples that applied these concepts in risk assessment to date are limited to individual chemicals, lacking an explicit treatment of internal biotransformation and of combined effects resulting from non-chemical stressors and chemical mixtures. Moving from linear AOPs to AOP networks is one step closer towards the idea of integrating both the exposome approach and the AOP concept (Fig. 1). We argue that the AOP concept can expand applications of the exposome beyond EWAS to establish mechanistic understanding of the causal linkages between chemical exposure and adverse effects. In turn, the exposome approach can help the AOP concept grow beyond single chemicals to include the effect of jointly acting mixtures that include both chemical and non-chemical stressors.

In addition, we may profit from the analogy of the environmental and human exposure via the food chain and other uptake pathways via air, water, dust, and consumer-products (Fig. 1). We consider experimental tools that can connect both research arenas to better understand well-conserved effects on the cellular level – emphasizing commonalities and differences between the anthroposphere and the ecosphere.

Here we first explore the history of the exposome and AOP concepts and then propose to use tools from systems chemistry and systems biology to integrate exposome and AOP concepts. We then conclude with recommendations for further research.

1.1. Defining relevant exposure

1.1.1. Exposome

In 2005, Wild coined the term “exposome” to describe the entirety of environmental exposure that, as a complement to the genome, may provide important clues for the understanding of chronic diseases. In his concept, the exposome encompasses lifetime environmental exposures that include lifestyle factors from the prenatal period onwards (Wild, 2005). From this view, an accurate assessment of a complete exposure history is required to understand the complex interplay with genetic susceptibility, since the majority of genetic alterations will contribute to population disease burden only in the presence of specific environmental exposures (Vineis et al., 2001; Wild, 2005).

Rappaport and Smith refined the approach through emphasizing the role of the chemistry in the organism, defining the exposome as the totality of human exposures from all exogenous and endogenous sources in the “internal chemical environment” (Rappaport and Smith, 2010). This latter definition takes into account that exposures are comprised not only of chemicals entering the body from the environment (e.g.,

air, food, water, dust), but also include compounds produced in the body by inflammation, (oxidative) stress, lipid peroxidation, infections, the microbiome, and other natural processes (Fig. 2). The internal chemical environment is highly dynamic during lifetime due to (environmental) external and internal factors and processes such as aging, infections, lifestyle, preexisting diseases etc. (Rappaport, 2011; Rappaport and Smith, 2010).

The above-mentioned definitions of the exposome have their specific merits depending on the angle from which exposure is viewed. “Bottom-up” strategies focus on pre-selected compounds or compound adducts following targeted hypotheses. By contrast, “top-down” strategies aim at measuring all chemicals or products of their downstream processing in a subject’s biospecimen, such as blood (Rappaport, 2011; Rappaport and Smith, 2010), as far as technically feasible. However, the full characterization of the exposome throughout the whole lifespan remains an outstanding challenge.

1.1.2. Eco-exposome

While the exposome was originally defined to characterize human exposures, the idea can certainly be adapted to consider ecosystem exposure. In 2012, the National Research Council (NRC) of the US National Academy of Sciences defined the “eco-exposome” as “the extension of exposure science from the point of contact between a stressor and receptor inward into the organism and outward to the general environment, including the ecosphere” (National Research Council (NRC), 2012). Similar to the extended definitions of the human exposome, the eco-exposome is described by both internal and external measures of exposure (Lioy and Smith, 2013). Naturally, the implementation of the exposome approach is specific for each biological species. However, conservation of targets for drugs and chemicals on the cellular level (Gunnarsson et al., 2008) might allow links to be found across species and between humans and ecosystems (LaLone et al., 2014; Rand-Weaver et al., 2013). The exposome narrative could establish an important link between human and ecosystem health by examining the effect of the totality of exposure from exogenous and endogenous sources over all levels of biological organization and complexity (Fig. 1). Although the eco-exposome adds another level of complexity, shared toxicity pathways and adaptive stress responses can be invoked to identify commonalities (Kramer et al., 2011), and variations in metabolic and functional traits may explain differences (Forbes and Galic, 2016).

There are examples where elements of exposome research have been applied in ecotoxicology, such as monitoring contaminants in whole organisms (Houde et al., 2011; Lana et al., 2014; Lehnert et al., 2016) or correlative studies associating functional health parameters with body burdens of organic pollutants in marine wildlife (Jin et al., 2015). Similarly to exposome research in human health, there has been a focus on exposure assessment without necessarily establishing a quantitative link to adverse effects. Hence, the challenges of, both, the human exposome and eco-exposome are similar and there is mutual benefit for generating mechanistic knowledge.

1.2. Application of the AOP concept in the context of exposome research

A major motivation for developing the AOP concept was to support the risk assessment of chemicals by providing mechanistic knowledge that would enable to link *in vitro* (Andersen et al., 2005), *in chemico* (Böhme et al., 2009) or *in silico* information (Rusyn and Daston, 2010), including computational biotransformation (Ji and Schüürmann, 2013) and structural alerts (Schüürmann et al., 2016), to toxicity *in vivo* (Berggren et al., 2015; Edwards et al., 2016). The AOP concept supports the validation of predictive models based on underlying molecular mechanisms. This would allow a more efficient use of high throughput screening (HTS) to prioritize a large number of compounds for detailed testing and/or reducing the number of animal experiments (Hartung et al., 2013b). Furthermore, within a regulatory framework of integrated

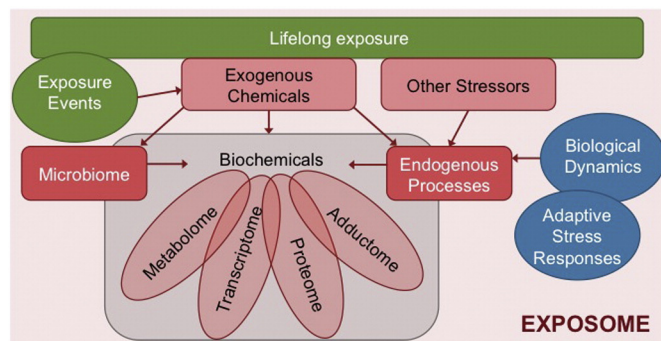


Fig. 2. Defining the exposome.

assessment, AOPs can support a new, more targeted safety testing regimen that is focused on the most probable and relevant hazards (Rovida et al., 2015; Tollefsen et al., 2014). Rather than apical endpoints typically assessed for regulatory purposes, subchronic or sublethal endpoints, as well as *in vitro* bioassays related to acute and chronic adverse effects, may be used for establishing AOPs. The AOP concept has the inherent promise to evolve from a tool for structuring knowledge or prioritization of testing to a quantitative predictive tool to relate exposure data to adverse outcomes (AO). However, challenges remain that concern for example the incorporation of toxicokinetics as important determinant of toxicity in AOPs and extrapolations to higher levels of biological organization and across species (Groh et al., 2015). Thus, substance or species-specific differences in the toxicokinetics and genetically based differences across species can influence single events or processes within an AOP cascade, in consequence augmenting or mitigating apical adverse effects.

1.2.1. How could the exposome approach and the AOP concept cross-fertilize each other?

Initially, the exposome and the AOP concepts evolved separately in the fields of human toxicology and ecotoxicology, respectively. The inclusion of biological response in exposure assessment is where AOPs integrate into the exposome; the consideration of endogenous and non-chemical stress as well as mixture effects is where the exposome can enrich the AOP concept. Per definition, the exposome is specific for individuals and integrates lifetime exposure while the AOP is conceptually focused on biological mechanisms and pathways. The temporal aspects of the exposome are at least partially implemented in the AOP concept. For instance, life-stage specificity or chronic toxicity would address effects that are related to long-term or potentially repeated exposure scenarios and biological responses.

The system-wide analysis of biological responses employing toxicogenomics represents an unbiased approach to detect chemical effects, integrating from MIEs along toxicity pathways (Berninger et al., 2014; Ellinger-Ziegelbauer et al., 2009). However, large numbers of signals compromise the response signatures (Vidal-Dorsch et al., 2016) and may lead to over-fitted associations with exposure characteristics. This calls for a reduction in data dimensionality through mechanistic reasoning and biomarker identification (Blauboer et al., 2012) to discriminate relevant signals from epi-phenomena and random responses (Hartung et al., 2012). In this respect the combination of AOP and exposome is revisiting previous approaches to apply biomarkers in epidemiology (Toniolo et al., 1997). A biomarker may represent a KE leading to adverse outcomes but may also represent events that are without

direct relevance to the adverse effect, e.g. representing a compensatory reaction. However, the AOP concept is stricter and clearer with respect to the relation of events that are directly related to adverse effects and hence, its combination with the exposome could improve the relation of exposome signals to impacts on health.

AOPs could further help to anchor system-wide responses to dominant modes or mechanisms of action (Ellison et al., 2016), and thus increase the confidence in a hypothesis by providing mechanistic plausibility (Braun et al., 2016).

1.2.2. Aggregate exposure pathways (AEPs), AOPs, and the exposome

The exposome may also be considered as the integration of AEPs and AOPs on (complex) mixtures if the source and pathways leading to the internal exposure are included (Fig. 3). The AEP concept has been developed complementary to the AOP describing the KEs from source via external exposure (including environmental and dietary exposure) to exposure in the organism, termed target site exposure (Teeguarden et al., 2016), which is the crucial step linked to the molecular initiating events of the AOP. Similar to AOPs, the AEPs help to organize exposure information from exogenous source to internal site of action, setting the stage for inferring chemical concentrations at the internal target site and informing about expected biological effects.

The chemicals in the exposome constitute not only the exogenous chemicals transported into the body and their metabolites, but also adducts with cellular constituents and other endogenous compounds, as well as signaling molecules formed as part of the pathway of toxicity (Kleensang et al., 2014) or adaptive stress responses (Simmons et al., 2009; Smirnova et al., 2015) (Fig. 3). The formation of chemical adducts can be considered as MIE, while endogenous chemicals (e.g. reactive oxygen species, nitric oxide, ATP, glutathione) formed or changed in their levels as part of the stress response belong to the KE. Aligning the chemicals in the exposome to the various steps of the AEP/AOP (Fig. 3) will help to establish a clearer *a priori* mechanistic link between the exposome and adverse health outcomes, with some endogenous chemicals of the exposome involved in defense against rather than in the development of the disease. However, the internal chemical response of stressed/perturbed organisms (e.g., due to an infection or an unhealthy lifestyle) may potentially lower the repair capacities, which may result in the next downstream event. These specific cases should be addressed in future studies.

1.2.3. Putative AOPs in exposome assessment

The AOP concept has been moving from the initial ideas of linear pathways to networks of pathways (Knapen et al., 2015), which

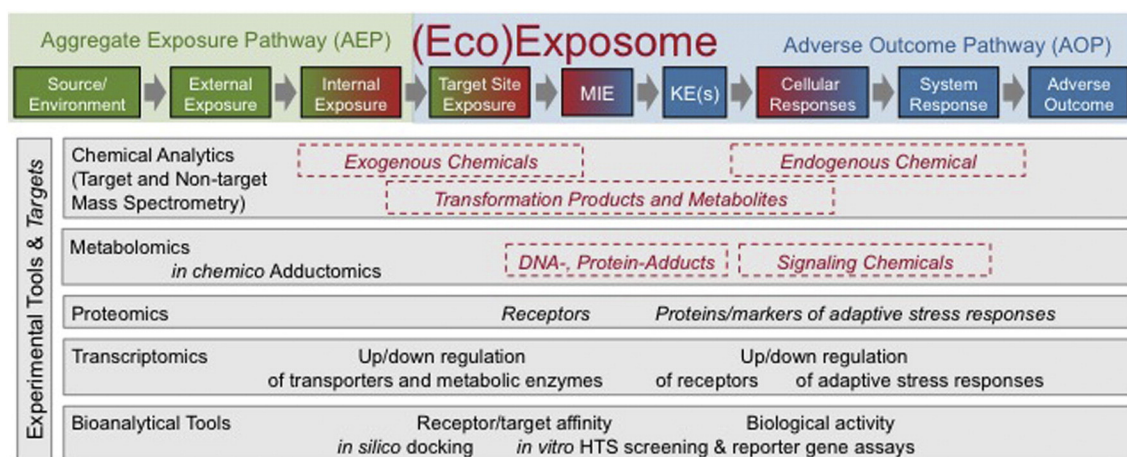


Fig. 3. Interface between the (eco)exposome (in red), the aggregate exposure pathway (AEP, green) and adverse outcome pathway (AOP, blue). The red dashed boxes represent chemical components of the exposome. The AEP/AOP concept allows one to disentangle key events and allocate them to steps from the source of exposure to adverse effects. The grey boxes indicate experimental methods to quantify the chemical components of the exposome and the biological components of the AOP. Figure partially adapted from Teeguarden et al. (2016). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

accommodates the idea of multiple causes for adverse effects. Some scientists criticize that AOPs are presently often incomplete. However, partial information on KEs and their relation even in case of weak evidence can provide initial steps to prioritize areas that require further investigation, and to identify the most relevant (internal) exposure situations. The AOP definitions and development are supported and guided by the OECD on a global level (Worth et al., 2014). Putative AOPs can be validated by the same principles that apply to AOPs in general, that is, their consistency with scientific literature and evidence for mechanistic links has to be demonstrated (Bell et al., 2016; Hartung et al., 2013a). Furthermore, putative and partially incomplete AOPs with high confidence relationships between KEs can be useful in specific applications, such as predicting an adverse outcome based on an easily tested KE (Perkins et al., 2015).

1.2.4. Grouping of exposures with converging AOPs

Given the large chemical variability, a full characterization of all possible MIEs and pathway-specific KEs remains one of the greatest challenges of future research. However, many AOPs converge at a higher level of biological organization and the AOP-based assessment could be conducted at more downstream KEs using cellular or organ responses. Examples include neuroactive pesticides or compounds disrupting the thyroid axis. Many pesticides act by interfering with specific steps in neural signal transduction that converge at the level of the cardiovascular system leading to a respiratory failure syndrome and finally death of the organism (Bradbury et al., 2008). Hence, characterization of behavioral responses could allow the integration of various mechanisms and to comprehensively describe exposure to neurotoxic compounds. Compounds disrupting the thyroid hormone system provide related examples in human toxicology. While various different MIEs are known, they finally converge at the intracellular or systemic thyroid hormone level (Murk et al., 2013). Hence, an integrative assessment could be based on test systems that target hormone levels in an organism, e.g., by assessment of compensatory responses to reduced thyroid hormone levels (Fetter et al., 2015).

1.2.5. Quantitative versus qualitative AOPs

Most AOPs are initially developed based on qualitative, mechanistic evidence without consideration of toxicokinetics, i.e. the uptake, distribution and metabolism/elimination of a compound in an organism. However, the cellular concentration is a major driver of the magnitude of the final adverse effect. In the case of, e.g., a limited uptake or rapid metabolism, adverse effects may be mitigated even for a high affinity to the molecular target (Patlewicz et al., 2013).

Quantitative AOPs (qAOP) build on approaches for toxicokinetic-toxicodynamic (TKTD) modeling (MacKay et al., 2013) as was shown very recently for a qAOP developed for effects of synthetic glucocorticoids in fish (Margiotta-Casaluci et al., 2016). If the AOP cannot be fully described, hazard rates may be established to obtain a quantitative link to the final adverse effect (Ashauer et al., 2015).

1.2.6. Networks of AOPs for mixtures

If bioassays can be mapped to KEs in AOPs of specific chemical domains, AOP networks might be used for assessing the complex mixtures of the exposome. Studies on extracts of environmental samples have demonstrated how complex chemical mixtures can be characterized using a battery of different mechanistic bioassays (Escher et al., 2014) or transcriptomic tools (Berninger et al., 2014). The levels of single chemicals will often fall below detection limits (both for chemical analysis and biological responses), but their combined exposure may nevertheless generate detectable biological responses (Altenburger et al., 2012; Silva et al., 2002). An AOP-based approach could help to interpret effects of mixtures without necessarily resolving each component, e.g., if a mixture of chemicals is extracted from biospecimens and the resulting extract is applied to cell-based bioassays (Altenburger et al., 2015). Bioanalytical assessment of these extracts with cell-based

bioassays may support the identification of the most relevant chemicals and toxicity pathways interfering with human and wildlife health, provided that the bioassays are anchored in a defined step of the AOP (Busch et al., 2016). Identification of such risk drivers could define new target chemicals in (bio)monitoring programs.

The requirements for AOPs to be used for mixtures do not deviate principally from those for AOPs for single compounds (Altenburger et al., 2015). However, similar adverse outcomes may be triggered by different MIEs that could jointly affect the same KE or more complex interactions could arise as is delineated by the complex role of low-dose mixtures in carcinogenesis (Goodson et al., 2015). Hence, experimental methods should focus on those KEs that can be expected to aggregate the bioactivity of different compounds that bind to different target sites but still converge into the same adverse outcome (Fig. 4A, Vogs and Altenburger, 2016).

Current understanding of mixture effects offers this reasoning as an explanation of why mixtures act according to concentration addition or independent action and may still show a combined effect where the individual components occur at sub-threshold concentrations (Altenburger and Greco, 2009). So far, the predictive power of mixture models has been demonstrated mainly for artificially designed mixtures and defined apical endpoints (e.g., growth, survival) (Kortenkamp et al., 2009). More complex and environmentally relevant mixtures have occasionally been evaluated using the concentration addition assumption (Tang et al., 2013). In the past, mixture analysis that used endpoints anticipated to strongly relate to an MIE, such as toxicogenomic responses, were often based on poorly designed studies and therefore remained inconclusive (Altenburger et al., 2012). For the extension of the principles of mixture toxicity to the AOP concept it is thus necessary to explore how AOPs originating from different MIEs may converge at the KE and/or AO level and whether AOPs can be formulated for complex mixtures that cannot be resolved to an individual mechanism of action. However, even when a mixture effect would appear to be linked to an individual AOP, this AOP may not allow one to trace back to a certain chemical or chemical class due to the large number of chemicals and complex interactions within the overall exposome. Still, such a diagnosis would be informative because it would establish a mechanistic link between exposure and adverse effects.

Furthermore, toxicodynamic models are at present not available for mixtures. They would be an asset to quantitatively model and predict adverse outcomes or diseases resulting from mixtures for which exposure information is limited or where quantitative knowledge is available only for a limited number of KEs (Fig. 4B). Such models would also open an avenue towards approaching temporal issues in adverse outcome assessment such as non-continuous or sequential exposures.

1.3. The role of systems biology and chemistry in exposome and AOP research

The methodological “glue” that links the exposome and AOPs consists, largely, of systems biological and chemical methods (Fig. 3). Systems biology is characterized by (i) an initial massive parallel experimental approach, aimed at determining one or more levels of molecular signatures (e.g., transcriptome, proteome, metabolome etc.); (ii) an iterative integration of experimental approaches and computational data analysis, and modeling, and (iii) the computational generation of experimentally testable hypotheses (Garcia-Reyero and Perkins, 2011; Hood et al., 2012; Ideker et al., 2001).

The complementary view on systems involving molecules with their compartmental partitioning and reaction networks is called systems chemistry. It addresses the organization of molecular feedback, amplification and information gain, and how these translate into the emergence of system-level properties (Ludlow and Otto, 2008; Nitschke, 2009; Whitesides, 2015). In the context of the exposome, systems chemistry may describe the interplay between exogenous and

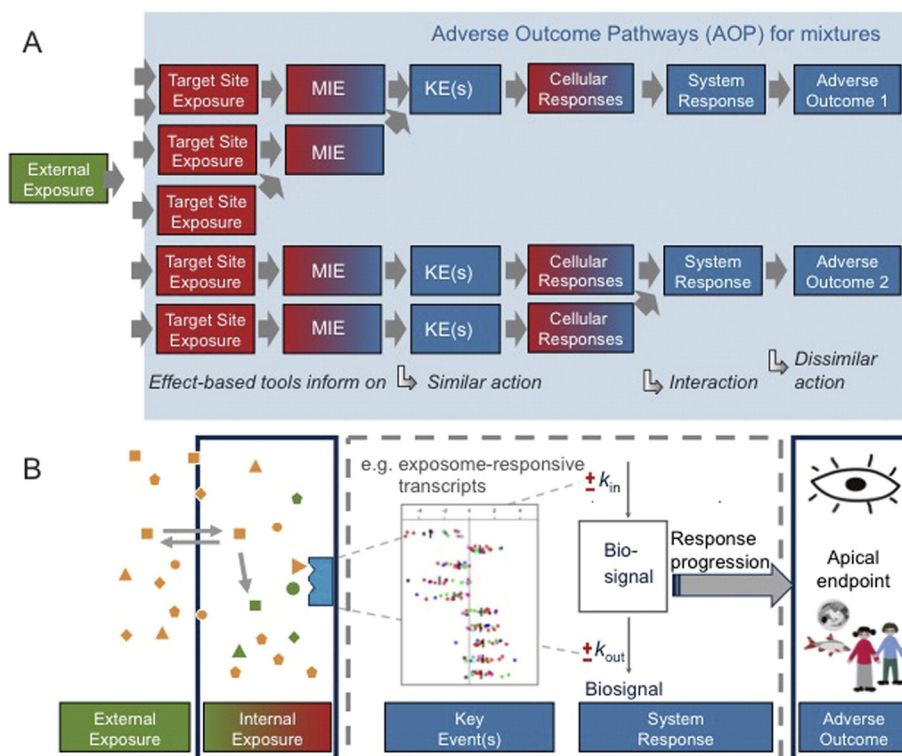


Fig. 4. (A) AOP for mixtures and (B) qAOP mixture modeling concept for similar action. Adapted with permission from Vogs, C. and Altenburger, R. (2016) Time-Dependent Effects in Algae for Chemicals with Different Adverse Outcome Pathways: A Novel Approach. *Environmental Science & Technology*, 50(14): 7770–7780. Copyright (2016) American Chemical Society.

endogenous compounds including their dependence on the spatially varying dose as well as on reactivity, time, and further system properties.

Since chemistry underpins biological processes both in organisms and in the environment, a way forward is to combine omics and analytics with computational tools. Interdisciplinary approaches and bioinformatics tools may serve to identify perturbations in organisms and in the environment, to define biomarkers of exposure and disease, and to integrate information from all relevant levels of organization (Smith et al., 2015). AOPs can be regarded as biological roadmaps along which chemistry mediates the development of toxicological effects. Identification and quantification of AOPs involve chemicals as both triggers and modulators of toxicity, and employ systems biology methods to reconstruct regulatory pathways, and to assess perturbations and rewiring in biological networks (Fig. 3).

1.3.1. What to measure?

The first step is to use advanced analytics for identifying and quantifying exogenous and endogenous chemicals (Fig. 3). EWAS are limited by the inherent definition of 'exposure-wide' that calls for untargeted analytical approaches (Patel, 2016). Despite enormous advancement in analytical methodologies in recent years, problems persist and only a very small number of the thousands of compounds detectable in a sample can actually be identified, leaving the largest fraction of chemicals at the level of a known accurate mass (or molecular formula) and retention time. Any improvements here rely strongly on a better assignment of likely structures for these peaks based on a prediction of fragmentation, ionization, or chromatographic retention times supported by more comprehensive mass spectra databases. In contrast to non-targeted analysis, targeted approaches require initial hypotheses regarding classes of analytes and thus cannot carry through on the promise of 'exposure-wide' detection. Currently, efforts are under way to develop automated workflows to analyze large analytical datasets, including multivariate statistical approaches dealing with patterns of

chemical signals in relationship to adverse outcomes without attempting identification and quantification of individual chemicals (Patel, 2016).

The characterization of the impact of external factors on the internal chemical environment calls for the use of omics and analytical techniques (Fig. 3). In recent years, substantial progress has been made in measuring small molecules and metabolites (metabolomics, Athersuch and Keun, 2015), DNA adducts (adductomics, Phillips et al., 2014; Rappaport et al., 2012) or large molecules, such as proteins and peptides (proteomics, Stallman Brown, 2012).

Transcriptomics – except emerging techniques to detect chemically modified RNA – does not identify products of external stress directly, but has proven valuable for characterizing responses to exposure at the pathway level. The transcriptome is easily assessed – also in cohort studies – since it can be applied even if only little material down to single cells is available. By combining transcriptomics with classifier analysis, more targeted endpoints can be developed (de Boer et al., 2015).

Also of interest is the assessment of environment-induced epigenetic changes, some of which have been shown to persist for prolonged periods of time (Bauer et al., 2016; Guida et al., 2015). Epigenomics may thus provide stable biomarkers for past exposures that are not detectable due to a limited sampling scheme by other omics layers. However, interpretation of epigenomic data remains challenging and functional assessment of epigenomic changes frequently requires integration with other omics data sets.

To assist in interpreting large-scale exposure data, the AOP concept may help to establish mechanistic links to external perturbation, for example by high-throughput screening of chemical interference with specific cellular toxicity pathways. Reporter gene assays based on nuclear receptors and transcription factors of cellular toxicity pathways have become popular tools not only to quantify exposure, but also to identify relevant steps of the AOP triggered by chemicals and their complex environmental mixtures (Fig. 3).

1.3.2. Where to measure?

In human exposomics, several biospecimens are appropriate for assessing the internal chemical environment. Urine can be sampled non-invasively, but favors the detection of water-soluble chemicals, including metabolites and conjugates. Measuring the blood exposome is a sensible approach compared to organ-specific samples (Rappaport et al., 2014). Blood transports chemicals to and from tissues and represents a reservoir of many endogenous and exogenous chemicals in the body at a given time (Nicholson et al., 2012). Blood samples are collected in most cohort studies, are therefore easily accessible, and sometimes a drop of blood is sufficient for biomonitoring studies (Mao and Wang, 2015).

In eco-exposomics, integrating external with internal exposure has practical but also theoretical implications: In small aquatic invertebrates or fish embryos, the whole organisms need to be extracted. Analysis of whole organisms is based on extracts from a poorly defined mixture of lipids, proteins, carbohydrates, and bodily fluids. For larger organisms with more complex exposure pathways, such as mammals, larger fish or birds, the exposome may be investigated in bodily fluids similar to the human exposome approach.

The AOP components involving MIEs and pathway responses in key processes, such as development, are often evolutionarily conserved between human and model organism. Cross-species comparison focused on conserved KEs allows for AOP anchoring, assuming that evolutionary conserved proteins may have conserved functions. Therefore identification of protein orthologs through sequence similarity or other methods might be helpful to infer susceptibility, particularly if an AOP has already been identified (Perkins et al., 2013). Differences between species that cannot be captured by this approach would be toxicokinetic differences, especially with respect to metabolism.

1.3.3. When to measure?

The human exposome is highly variable and dynamic throughout the human lifespan. The time-dependence of the exposome poses a challenge for the integration with the AOP concept. AOPs so far do not adequately reflect when KEs or KE relationships are only valid during a certain life stage or developmental phase.

Routine measurements, in particular during critical life stages, such as fetal development (cord blood analysis), early childhood, and puberty, are key to establishing a personalized picture of specific individuals' exposures (Rappaport and Smith, 2010). Time-sequenced information on individual exposome at different time points prior to disease onset would be of immense relevance for identifying environmental causes of disease.

Human life-long exposure may leave its traces in the exposome, while a retrospective analysis of external exposure along all uptake pathways is impossible. For small and short-lived aquatic organisms, however, intelligent environmental sampling regimes yield time-integrated and peak exposures, bringing us close to the ideal of an assessment of life-long exposure to external chemicals.

1.3.4. From exposome to adverse outcomes

Systems biology and chemistry tools may be linked to prioritize compounds for chemical analytics by a tiered approach: Starting from a population cohort with a nested case-control study – “meet in the middle” approach that combines bottom-up and top-down approaches (Vineis et al., 2013) – omics data from epigenomics, proteomics or transcriptomics are generated and used to identify pathways/networks potentially affected by exposures and subsequently driving disease risk. For these pathways, *in silico* exploitation of (toxicological) databases and chemical bioactivity from HTS, reporter gene assays and docking studies will yield pathway-associated exogenous and endogenous chemicals (Fig. 3).

Tox 21 and ToxCast have demonstrated the practicality of combining (hundreds of) HTS screens with *in vitro* bioassays for more than 8000 single compounds (Tice et al., 2013). While the current focus is *in vitro*

to *in vivo* extrapolation using toxicokinetic models (Phillips et al., 2016; Wetmore et al., 2013), these *in vitro* bioassays can also be applied to monitor unknown mixtures in environmental samples from water (Escher et al., 2014) to biota (Jin et al., 2015) and human specimens. In this regard, *in vitro* assays may be useful to capture endogenous exposures and changes in internal of stress. Methods to link cause with effect, such as effect-directed analysis, are well established for the analysis of the external exposure in water (Brack et al., 2016) or sediment (Brack et al., 2005) and are increasingly applied to mammals and fish bio-fluids and tissues. Mode-of-action-specific bioassays have been used together with fractionation approaches and untargeted analysis to identify drivers of potential adverse effects in wildlife (Houtman et al., 2007; Simon et al., 2013).

Certainly, not all prerequisites for such an approach are currently available. The link between pathway perturbation and MIE may not be traceable for a particular omics data set, specifically when lacking data for the appropriate time points. However, the increasing capability to acquire multi-omic data sets at several time-points in population cohorts and environmental populations, combined with the use of defined *in vitro* HTS tools (Tufi et al., 2016), may alleviate this issue.

1.3.5. Systems biology challenges in exposome research

Depending on the kinetics between initial exposure, MIEs and subsequent adaptations, different levels of molecular responses may require analysis (Yugi et al., 2016). Regarding prenatal exposure, long-term health effects might be accessible through epigenetics, whereas the metabolome level appears best for immediate effects induced by environmental chemicals.

Cross-omics data integration is particularly relevant for AOP construction: the multi-scale nature of AOPs calls for integrating more than one omics layer. Single-cell pathway level effects will most likely be studied using respective proteomics or transcriptomics, whereas the organismal and population effects might be more easily captured using serum metabolomics. Currently available approaches reach their limits when the need arises to capture complex non-linear relationships between different omics data sets such as feedback loops.

Omics data used for network inference and pathway analysis should ideally be assessed in a time-resolved manner (Bar-Joseph et al., 2012). Most methods used for network reconstruction do not model time explicitly (Hempel et al., 2011) and more recent methods that do may demand further analysis to fully assess their potential (Le Novère, 2015).

2. Conclusions

2.1. How the AOP concept enhances understanding the exposome and its impact on adverse outcome

An inspection of publications on AOP and exposome from the past decade indicates that the concepts are widely accepted and intensively discussed but that examples of practical applications remain scarce. It can be anticipated that this will change in the near future provided that application of the concepts in research are intensified. While both concepts are in the midst of development, their integration might lead to synergy, which will be promoted by early harmonization of data collection and terminology.

The exposome approach and the AOP concept are essentially orthogonal with the AOPs covering single pathways over the entire effect chain while the exposome covers a multitude of pathways, with the exposure in principle integrated over the lifetime but in practice as cross sectional studies at a fixed time over a population. Thus, the crossing points of AOP and exposome will need to be expanded in both dimensions.

To further explore the utility and benefit of future application of AOPs in exposome assessment the following research topics need to be strengthened:

- 1) More AOPs should be developed and deposited in central databases (e.g., <https://aopwiki.org/>). With every new AOP developed the capacity of exposome assessment based on biological responses will increase significantly.
- 2) Evolutionarily conserved cellular toxicity pathways may serve as common denominators for integrated effect assessments. We advocate the use of KEs across species for a systems biology-assisted approach to exposome assessment.
- 3) Chronic exposure (both in terms of exposure duration and delay of effects) represents the typical environmental situation and is most relevant for human health. AOPs for chronic toxicity are at present not well-described (Groh et al., 2015). However, one example for a chronic AOP is described in the AOP wiki, i.e. “Chronic binding of antagonist to N-methyl-D-aspartate receptors (NMDARs) during brain development induces impairment of learning and memory abilities” (<https://aopwiki.org/>). Furthermore, many other endpoints, such as reproductive dysfunction originating from endocrine disruption are of relevance only in case of a chronic exposure since short-term exposures may not lead to significant population decline. Hence, an AOP-based exposome assessment should target chronic endpoints and disease outcomes.
- 4) The research question is an important driver of the type of exposome/AOP research, such as the identification of the main chemical risk drivers in relation to mode of action, the complex multifactorial influences of mixtures or identification of a threshold level that can explain adverse effects.

2.2. Mixtures

Existing mixture models have to be adapted and tested with regard to the AOP and exposome concepts. How established mixture toxicity models can be applied to different KEs that lead to the same adverse outcome remains a key question for future research.

2.3. Exposome characterization as a driver for risk assessment

Characterizing the exposome via untargeted measurement of an internal chemical environment promotes data-driven discoveries of causal factors for human diseases or effects on the ecosystem. To identify the sources and to develop prevention strategies, the main exposures have to be characterized and validated by targeted techniques. Moreover, the discrimination of different exposures also provides the basis for hypothesis-driven research to promote mechanistic understanding. A promising approach to further develop such a mechanistic understanding will be the use of the AOP concept as long as it is evolving further from a linear pathway analysis to a tool to organize the complex networks of toxicity pathways.

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References

- Altenburger, R., Greco, W.R., 2009. Extrapolation concepts for dealing with multiple contamination in environmental risk assessment. *Integr. Environ. Assess. Manag.* 5, 62–68.
- Altenburger, R., Scholz, S., Schmitt-Jansen, M., Busch, W., Escher, B.I., 2012. Mixture toxicity revisited from a toxicogenomic perspective. *Environ. Sci. Technol.* 46, 2508–2522.
- Altenburger, R., Ait-Aissa, S., Antczak, P., Backhaus, T., Barcelo, D., Seiler, T.-B., Brion, F., Busch, W., Chipman, K., de Alda, M.L., de Aragao Umbuzeiro, G., Escher, B.I., Falciani, F., Faust, M., Focks, A., Hilscherova, K., Hollender, J., Hollert, H., Jager, F., Jahnke, A., Kortenkamp, A., Krauss, M., Lemkine, G.F., Munthe, J., Neumann, S., Schymanski, E.L., Scrimshaw, M., Segner, H., Slobodnik, J., Smedes, F., Kughathas, S., Teodorovic, I., Tindall, A.J., Tollefsen, K.E., Walz, K.-H., Williams, T.D., Van den Brink, P.J., van Gils, J., Vrana, B., Zhang, X., Brack, W., 2015. Future water quality monitoring - adapting tools to deal with mixtures of pollutants in water resource management. *Sci. Total Environ.* 512–513, 540–551.
- Andersen, M.E., Dennison, J.E., Thomas, R.S., Conolly, R.B., 2005. New directions in incidence-dose modeling. *Trends Biotechnol.* 23, 122–127.
- Ankley, G.T., Bennett, R.S., Erickson, R.J., Hoff, D.J., Hornung, M.W., Johnson, R.D., Mount, D.R., Nichols, J.W., Russom, C.L., Schmieder, P.K., Serrano, J.A., Tietge, J.E., Villeneuve, D.L., 2010. Adverse outcome pathways: a conceptual framework to support ecotoxicology research and risk assessment. *Environ. Toxicol. Chem.* 29, 730–741.
- Ashauer, R., O'Connor, I., Hintermeister, A., Escher, B.I., 2015. Death dilemma and organ-ism recovery in ecotoxicology. *Environ. Sci. Technol.* 49, 10136–10146.
- Athersuch, T.J., Keun, H.C., 2015. Metabolic profiling in human exposome studies. *Mutagenesis* 30, 755–762.
- Bar-Joseph, Z., Gitter, A., Simon, I., 2012. STUDY DESIGNS studying and modelling dynamic biological processes using time-series gene expression data. *Nat. Rev. Genet.* 13, 552–564.
- Bauer, T., Trump, S., Ishaque, N., Thurmann, L., Gu, L., Bauer, M., Bieg, M., Gu, Z., Weichenhan, D., Mallm, J.-P., Roder, S., Herberich, G., Takada, E., Mucke, O., Winter, M., Junge, K.M., Grutzmann, K., Rolle-Kampczyk, U., Wang, Q., Lawerenz, C., Borte, M., Polte, T., Schlesner, M., Schanne, M., Wiemann, S., Georg, C., Stunnenberg, H.G., Plass, C., Rippe, K., Mizuguchi, J., Herrmann, C., Eils, R., Lehmann, I., 2016. Environment-induced epigenetic reprogramming in genomic regulatory elements in smoking mothers and their children. *Mol. Syst. Biol.* 12, 861.
- Bell, S.M., Angrish, M.M., Wood, C.E., Edwards, S.W., 2016. Integrating publicly available data to generate computationally predicted adverse outcome pathways for fatty liver. *Toxicol. Sci.* 150, 510–520.
- Berggren, E., Amcoff, P., Benigni, R., Blackburn, K., Carney, E., Cronin, M., Deluyker, H., Gautier, F., Judson, R.S., Kass, G.E.N., Keller, D., Knight, D., Lilienblum, W., Mahony, C., Rusyn, I., Schultz, T., Schwarz, M., Schüürmann, G., White, A., Burton, J., Lostia, A.M., Munn, S., Worth, A., 2015. Chemical safety assessment using read-across: assessing the use of novel testing methods to strengthen the evidence base for decision making. *Environ. Health Perspect.* 123, 1232–1240.
- Berninger, J.P., Martinovic-Weigelt, D., Garcia-Reyero, N., Escalon, L., Perkins, E.J., Ankley, G.T., Villeneuve, D.L., 2014. Using transcriptomic tools to evaluate biological effects across effluent gradients at a diverse set of study sites in Minnesota, USA. *Environ. Sci. Technol.* 48, 2404–2412.
- Betts, K.S., 2013. Tox21 to date steps toward modernizing human hazard characterization. *Environ. Health Perspect.* 121, A228.
- Blauboer, B.J., Boekelheide, K., Clewell, H.J., Daneshian, M., Dingemans, M.M., Goldberg, A.M., Heneweer, M., Jaworska, J., Kramer, N.I., Leist, M., 2012. The use of biomarkers of toxicity for integrating *in vitro* hazard estimates into risk assessment for humans. *ALTEX* 29, 411–425.
- Böhme, A., Thäns, D., Paschke, A., Schüürmann, G., 2009. Kinetic glutathione chemoassay to quantify thiol reactivity of organic electrophiles-application to alpha,beta-unsaturated ketones, acrylates, and propiolates. *Chem. Res. Toxicol.* 22, 742–750.
- Brack, W., Schirmer, K., Erdinger, L., Hollert, H., 2005. Effect-directed analysis of mutagens and ethoxyresorufin-O-deethylase inducers in aquatic sediments. *Environ. Toxicol. Chem.* 24, 2445–2458.
- Brack, W., Ait-Aissa, S., Burgess, R.M., Busch, W., Creusot, N., Di Paolo, C., Escher, B.I., Hewitt, L.M., Hilscherova, K., Hollender, J., Hollert, H., Jonker, W., Kool, J., Lamoree, M., Muschket, M., Neumann, S., Rostkowski, P., Ruttkies, C., Schollee, J., Schymanski, E.L., Schulze, T., Seiler, T.-B., Tindall, A.J., Umbuzeiro, G.D.A., Vrana, B., Krauss, M., 2016. Effect-directed analysis supporting monitoring of aquatic environments - an in-depth overview. *Sci. Total Environ.* 544, 1073–1118.
- Bradbury, S.P., Carlson, R.W., Henry, T.R., Padilla, S., Cowden, J., 2008. Toxic responses of the fish nervous system. *The Toxicology Of Fishes*. 417–455.
- Braun, J.M., Gennings, C., Hauser, R., Webster, T.F., 2016. What can epidemiological studies tell us about the impact of chemical mixtures on human health? *Environ. Health Perspect.* 124, A6–A9.
- Busch, W., Schmidt, S., Kühne, R., Schulze, T., Krauss, M., Altenburger, R., 2016. Micro-pollutants in European rivers: a mode of action survey to support the development of effect-based tools for water monitoring. *Environ. Toxicol. Chem.* 35, 1887–1899.
- de Boer, T.E., Janssens, T.K.S., Legler, J., van Straalen, N.M., Roelofs, D., 2015. Combined transcriptomics analysis for classification of adverse effects as a potential end point in effect based screening. *Environ. Sci. Technol.* 49, 14274–14281.
- Edwards, S.W., Tan, Y.-M., Villeneuve, D.L., Meek, M., McQueen, C.A., 2016. Adverse outcome pathways—organizing toxicological information to improve decision making. *J. Pharmacol. Exp. Ther.* 356, 170–181.

- Ellinger-Ziegelbauer, H., Aubrecht, J., Kleinjans, J.C., Ahr, H.-J., 2009. Application of toxicogenomics to study mechanisms of genotoxicity and carcinogenicity. *Toxicol. Lett.* 186, 36–44.
- Ellison, C.M., Piechota, P., Madden, J.C., Enoch, S.J., Cronin, M.T.D., 2016. Adverse outcome pathway (AOP) informed modeling of aquatic toxicology: QSARs, read-across, and interspecies verification of modes of action. *Environ. Sci. Technol.* 50, 3995–4007.
- Escher, B., Allinson, M., Altenburger, R., Bain, P., Balaguer, B., Busch, W., Crago, J., Humpage, A., Denslow, N., Dopp, E., Hilscherova, K., Kumar, A., Grimaldi, M., Jayasinghe, B., Jarosova, B., Jia, A., Makarov, S., Maruya, K., Medvedev, A., Mehinto, A., Mendez, J., Poulsen, A., Prochazka, E., Richard, J., Schifferli, A., Schlenk, D., Scholz, S., Shiraishi, F., Snyder, S., Su, G., Tang, J., van der Burg, B., van der Linden, S., Werner, I., Westerheide, S., Wong, C., Yang, M., Yeung, B., Zhang, X., Leusch, F., 2014. Benchmarking organic micropollutants in wastewater, recycled water and drinking water with *in vitro* bioassays. *Environ. Sci. Technol.* 48, 1940–1956.
- Fetter, E., Baldauf, L., Da Fonte, D.F., Ortmann, J., Scholz, S., 2015. Comparative analysis of goitrogenic effects of phenylthiourea and methimazole in zebrafish embryos. *Reprod. Toxicol.* 57, 10–20.
- Forbes, V.E., Galic, N., 2016. Next-generation ecological risk assessment: predicting risk from molecular initiation to ecosystem service delivery. *Environ. Int.* 91, 215–219.
- Garcia-Reyero, N., Perkins, E.J., 2011. Systems biology: leading the revolution in ecotoxicology. *Environ. Toxicol. Chem.* 30, 265–273.
- Goodson III, W.H., Lowe, L., Carpenter, D.O., Gilbertson, M., Ali, A.M., de Cerain Salsamendi, A.L., Lasfar, A., Carnero, A., Azqueta, A., Amedei, A., Charles, A.K., Collins, A.R., Ward, A., Salzberg, A.C., Colacci, A., Olsen, A.-K., Berg, A., Barclay, B.J., Zhou, B.P., Blanco-Aparicio, C., Baglole, C.J., Dong, C., Mondello, C., Hsu, C.-W., Naus, C.C., Yedjou, C., Curran, C.S., Laird, D.W., Koch, D.C., Carlin, D.J., Felsner, D.W., Roy, D., Brown, D.G., Ratovitski, E., Ryan, E.P., Corsini, E., Rojas, E., Moon, E.-Y., Laconi, E., Marongiu, F., Al-Mulla, F., Chiaradonna, F., Darroudi, F., Martin, F.L., Van Schooten, F.J., Goldberg, G.S., Wagemaker, G., Nangami, G., Calaf, G.M., Williams, G., Wolf, G.T., Koppen, G., Brunborg, G., Lyster, H.K., Krishnan, H., Ab Hamid, H., Yasaei, H., Sone, H., Kondoh, H., Salem, H.K., Hsu, H.-Y., Park, H.H., Koturbash, I., Miousse, I.R., Scovassi, A.I., Klauin, J.E., Vondracek, J., Raju, J., Roman, J., Wise Sr., J.P., Whitfield, J.R., Woodrick, J., Christopher, J.A., Ochieng, J., Fernandez Martinez-Leal, J., Weisz, J., Kravchenko, J., Sun, J., Prudhomme, K.R., Narayanan, K.B., Cohen-Solal, K.A., Moorwood, K., Gonzalez, L., Soucek, L., Jian, L., D'Abronzio, L.S., Lin, L.-T., Li, L., Gulliver, L., McCawley, L.J., Memeo, L., Vermeulen, L., Leyns, L., Zhang, L., Valverde, M., Khatami, M., Romano, M.F., Chapellier, M., Williams, M.A., Wade, M., Manjili, M.H., Leonart, M., Xia, M., Gonzalez, M.J., Karamouz, M.V., Kirsch-Volders, M., Zaccari, M., Kuemmerle, N.B., Singh, N., Cruickshanks, N., Kleinstreuer, N., van Larebeke, N., Ahmed, N., Ogunkua, O., Krishnakumar, P.K., Vadgama, P., Marignani, P.A., Ghosh, P.M., Ostrosky-Wegman, P., Thompson, P., Dent, P., Heneberg, P., Darbre, P., Leung, P.S., Nangia-Makker, P., Cheng, Q., Robey, R.B., Al-Temaimi, R., Roy, R., Andrade-Vieira, R., Sinha, R.K., Mehta, R., Vento, R., Di Fiore, R., Ponce-Cusi, R., Dornetshuber-Fleiss, R., Nahta, R., Castellino, R.C., Palorini, R., Abd Hamid, R., Langie, S.A.S., Eltom, S., Brooks, S.A., Ryeom, S., Wise, S.S., Bay, S.N., Harris, S.A., Papagerakis, S., Romano, S., Pavanello, S., Eriksson, S., Forte, S., Casey, S.C., Luanpitpong, S., Lee, T.-J., Otsuki, T., Chen, T., Massfelder, T., Sanderson, T., Guarnieri, T., Hultman, T., Dormoy, V., Odeiro-Marah, V., Sabbiseti, V., Maguer-Satta, V., Rathmell, W.K., Engstrom, W., Decker, W.K., Bisson, W.H., Rojanasakul, Y., Luqmani, Y., Chen, Z., Hu, Z., 2015. Assessing the carcinogenic potential of low-dose exposures to chemical mixtures in the environment: the challenge ahead. *Carcinogenesis* 36, S254–S296.
- Groh, K.J., Carvalho, R.N., Chipman, J.K., Denslow, N.D., Halder, M., Murphy, C.A., Roelofs, D., Rolaki, A., Schirmer, K., Watanabe, K.H., 2015. Development and application of the adverse outcome pathway framework for understanding and predicting chronic toxicity: I. Challenges and research needs in ecotoxicology. *Chemosphere* 120, 764–777.
- Guida, F., Sandanger, T.M., Castagne, R., Campanella, G., Polidoro, S., Palli, D., Krogh, V., Tumino, R., Sacerdote, C., Panico, S., Severi, G., Kyrtopoulos, S.A., Georgiadis, P., Vermeulen, R.C.H., Lund, E., Vineis, P., Chadeau-Hyam, M., 2015. Dynamics of smoking-induced genome-wide methylation changes with time since smoking cessation. *Hum. Mol. Genet.* 24, 2349–2359.
- Gunnarsson, L., Jauhiainen, A., Kristiansson, E., Nerman, O., Larsson, D.G.J., 2008. Evolutionary conservation of human drug targets in organisms used for environmental risk assessments. *Environ. Sci. Technol.* 42, 5807–5813.
- Hartung, T., van Vliet, E., Jaworska, J., Bonilla, L., Skinner, N., Thomas, R., 2012. Systems toxicology. *ALTEX* 29, 119–128.
- Hartung, T., Hoffmann, S., Stephens, M., 2013a. Food for thought ... mechanistic validation. *ALTEX* 30, 119–130.
- Hartung, T., Luechtefeld, T., Maertens, A., Kleensang, A., 2013b. Food for thought ... integrated testing strategies for safety assessments. *ALTEX* 30, 3–18.
- Hempel, S., Koseska, A., Nikoloski, Z., Kurths, J., 2011. Unraveling Gene Regulatory Networks From Time-Resolved Gene Expression Data - A Measures Comparison Study. 12. *BMC Bioinformatics*, p. 292.
- Hood, L., Balling, R., Auffray, C., 2012. Revolutionizing medicine in the 21st century through systems approaches. *Biotechnol. J.* 7, 992–1001.
- Houde, M., De Silva, A.O., Muir, D.C.G., Letcher, R.J., 2011. Monitoring of perfluorinated compounds in aquatic biota: an updated review PFCs in aquatic biota. *Environ. Sci. Technol.* 45, 7962–7973.
- Houtman, C.J., Booi, P., van der Valk, K.M., van Bodegom, P.M., van den Ende, F., Gerritsen, A.A.M., Lamoree, M.H., Legler, J., Brouwer, A., 2007. Biomonitoring of estrogenic exposure and identification of responsible compounds in bream from Dutch surface waters. *Environ. Toxicol. Chem.* 26, 898–907.
- Ideker, T., Galitski, T., Hood, L., 2001. A new approach to decoding life: systems biology. *Annu. Rev. Genomics Hum. Genet.* 2, 343–372.
- Ji, L., Schüürmann, G., 2013. Model and mechanism: N-hydroxylation of primary aromatic amines by cytochrome P450. *Angew. Chem. Int. Ed.* 52, 744–748.
- Jin, L., Gaus, C., Escher, B., 2015. Bioanalytical approaches to understanding toxicological implications of mixtures of persistent organic pollutants in marine wildlife. In: Zheng, E. (Ed.), *Comprehensive Analytical Chemistry Persistent Organic Pollutants (POPs): Analytical Techniques, Environmental Fate and Biological Effects*. Elsevier.
- Kleensang, A., Maertens, A., Rosenberg, M., Fitzpatrick, S., Lamb, J., Auerbach, S., Brennan, R., Crofton, K.M., Gordon, B., Fornace Jr., A.J., Gaido, K., Gerhold, D., Haw, R., Henney, A., Ma'ayan, A., McBride, M., Monti, S., Ochs, M.F., Pandey, A., Sharan, R., Stierum, R., Tugendreich, S., Willett, C., Wittwehr, C., Xia, J., Patton, G.W., Arvidson, K., Bouhifd, M., Hogberg, H.T., Luechtefeld, T., Smirnova, L., Zhao, L., Adeleye, Y., Kanehisa, M., Carmichael, P., Andersen, M.E., Hartung, T., 2014. Pathways of toxicity. *ALTEX* 31, 53–61.
- Knapen, D., Vergauwen, L., Villeneuve, D.L., Ankley, G.T., 2015. The potential of AOP networks for reproductive and developmental toxicity assay development. *Reprod. Toxicol.* 56, 52–55.
- Kortenkamp, A., Backhaus, T., Faust, M., 2009. State Of The Art Report On Mixture Toxicity. European Commission (070307/2007/485103/ETU/D.1; 2009).
- Kramer, V.J., Etterson, M.A., Hecker, M., Murphy, C.A., Roesijadi, G., Spade, D.J., Spromberg, J.A., Wang, M., Ankley, G.T., 2011. Adverse outcome pathways and ecological risk assessment bridging to population-level effects. *Environ. Toxicol. Chem.* 30, 64–76.
- LaLone, C.A., Berninger, J.P., Villeneuve, D.L., Ankley, G.T., 2014. Leveraging existing data for prioritization of the ecological risks of human and veterinary pharmaceuticals to aquatic organisms. *Philos. Trans. R. Soc. B* 369, 20140022.
- Lana, N.B., Berton, P., Covaci, A., Ciocco, N.F., Barrera-Oro, E., Atencio, A., Altamirano, J.C., 2014. Fingerprint of persistent organic pollutants in tissues of Antarctic notothenioid fish. *Sci. Total Environ.* 499, 89–98.
- Le Novère, N., 2015. Quantitative and logic modelling of molecular and gene networks. *Nat. Rev. Genet.* 16, 146–158.
- Lehnert, K., Ronnenberg, K., Weijls, L., Covaci, A., Das, K., Hellwig, V., Siebert, U., 2016. Xenobiotic and immune-relevant molecular biomarkers in harbor seals as proxies for pollutant burden and effects. *Arch. Environ. Contam. Toxicol.* 70, 106–120.
- Lioy, P.J., Smith, K.R., 2013. A discussion of exposure science in the 21st century: a vision and a strategy. *Environ. Health Perspect.* 121, 405–409.
- Ludlow, R.F., Otto, S., 2008. Systems chemistry. *Chem. Soc. Rev.* 37, 101–108.
- MacKay, C., Davies, M., Summerfield, V., Maxwell, G., 2013. From pathways to people: applying the adverse outcome pathway (AOP) for skin sensitization to risk assessment. *ALTEX* 30, 473–486.
- Mao, P., Wang, D., 2015. Biomonitoring of Perfluorinated compounds in a drop of blood. *Environ. Sci. Technol.* 49, 6808–6814.
- Margiotta-Casaluci, L., Owen, S.F., Huerta, B., Rodriguez-Mozaz, S., Kugathas, S., Barcelo, D., Rand-Weaver, M., Sumpter, J.P., 2016. Internal exposure dynamics drive the adverse outcome pathways of synthetic glucocorticoids in fish. *Sci. Rep.* <http://dx.doi.org/10.1038/srep21978>.
- Miller, G.W., Jones, D.P., 2014. The nature of nurture: refining the definition of the exposome. *Toxicol. Sci.* 137, 1–2.
- Murk, A.J., Rijntjes, E., Blaauw, B.J., Clewell, R., Crofton, K.M., Dingemans, M.M., Furlow, J.D., Kavlock, R., Kohrle, J., Opitz, R., Traas, T., Visser, T.J., Xia, M., Gutleb, A.C., 2013. Mechanism-based testing strategy using *in vitro* approaches for identification of thyroid hormone disrupting chemicals. *Toxicol. In Vitro* 27, 1320–1346.
- National Research Council, 2007. *Toxicity Testing In The 21st Century: A Vision And A Strategy*. National Academies Press, Washington DC.
- National Research Council (NRC), 2012. *Exposure Science In The 21st Century: A Vision And A Strategy*. National Academic Press, Washington, DC.
- Nicholson, J.K., Holmes, E., Kinross, J.M., Darzi, A.W., Takats, Z., Lindon, J.C., 2012. Metabolic phenotyping in clinical and surgical environments. *Nature* 491, 384–392.
- Nitschke, J.R., 2009. Systems chemistry: molecular networks come of age. *Nature* 462, 736–738.
- Patel, C.J., 2016. Analytical complexity in detection of gene variant-by-environment exposure interactions in high-throughput genomic and exposomic research. *Curr. Environ. Health Rep.* 3, 64–72.
- Patlewicz, G., Simon, T., Goyak, K., Phillips, R.D., Rowlands, J.C., Seidel, S.D., Becker, R.A., 2013. Use and validation of HT/HC assays to support 21st century toxicity evaluations. *Regul. Toxicol. Pharmacol.* 65, 259–268.
- Perkins, E.J., Ankley, G.T., Crofton, K.M., Garcia-Reyero, N., LaLone, C.A., Johnson, M.S., Tietge, J.E., Villeneuve, D.L., 2013. Current perspectives on the use of alternative species in human health and ecological hazard assessments. *Environ. Health Perspect.* 121, 1002–1010.
- Perkins, E.J., Antczak, P., Burgoon, L., Falciani, F., Garcia-Reyero, N., Gutsell, S., Hodges, G., Kienzler, A., Knapen, D., McBride, M., Willett, C., 2015. Adverse outcome pathways for regulatory applications: examination of four case studies with different degrees of completeness and scientific confidence. *Toxicol. Sci.* 148, 14–25.
- Phillips, D.H., Preston, G., Sozeri, O., 2014. Adductomics - measurement of DNA and protein adducts in human tissues as an approach to defining the exposome. *Mutagenesis* 29, 502.
- Phillips, M.B., Leonard, J.A., Grulke, C.M., Chang, D.T., Edwards, S.W., Brooks, R., Goldsmith, M.-R., El-Masri, H., Tan, Y.-M., 2016. A workflow to investigate exposure and pharmacokinetic influences on high-throughput *in vitro* chemical screening based on adverse outcome pathways. *Environ. Health Perspect.* 124, 53–60.
- Rand-Weaver, M., Margiotta-Casaluci, L., Patel, A., Panter, G.H., Owen, S.F., Sumpter, J.P., 2013. The read-across hypothesis and environmental risk assessment of pharmaceuticals. *Environ. Sci. Technol.* 47, 11384–11395.
- Rappaport, S.M., 2011. Implications of the exposome for exposure science. *J. Expo. Sci. Environ. Epidemiol.* 21, 5–9.
- Rappaport, S.M., 2012. Biomarkers intersect with the exposome. *Biomarkers* 17, 483–489.
- Rappaport, S.M., Smith, M.T., 2010. Environment and disease risks. *Science* 330, 460–461.
- Rappaport, S.M., Li, H., Grigoryan, H., Funk, W.E., Williams, E.R., 2012. Adductomics: characterizing exposures to reactive electrophiles. *Toxicol. Lett.* 213, 83–90.

- Rappaport, S.M., Barupal, D.K., Wishart, D., Vineis, P., Scalbert, A., 2014. The blood exposome and its role in discovering causes of disease. *Environ. Health Perspect.* 122, 769–774.
- Rovida, C., Alepee, N., Api, A.M., Basketter, D.A., Bois, F.Y., Caloni, F., Corsini, E., Daneshian, M., Eskes, C., Ezendam, J., Fuchs, H., Hayden, P., Hegele-Hartung, C., Hoffmann, S., Hubesch, B., Jacobs, M.N., Jaworska, J., Kleensang, A., Kleinstreuer, N., Lalko, J., Landsiedel, R., Lebreux, F., Luechtefeld, T., Locatelli, M., Mehling, A., Natsch, A., Pitchford, J.W., Prater, D., Prieto, P., Schepky, A., Schüürmann, G., Smirnova, L., Toole, C., van Vliet, E., Weisensee, D., Hartung, T., 2015. Integrated testing strategies (ITS) for safety assessment. *ALTEX* 32, 25–40.
- Rusyn, I., Daston, G.P., 2010. Computational toxicology: realizing the promise of the toxicity testing in the 21st century. *Environ. Health Perspect.* 118, 1047–1050.
- Schüürmann, G., Ebert, R.-U., Tluczkiwicz, I., Escher, S.E., Kühne, R., 2016. Inhalation threshold of toxicological concern (TTC) – structural alerts discriminate high from low repeated-dose inhalation toxicity. *Environ. Int.* 88, 123–132.
- Silva, E., Rajapakse, N., Kortenkamp, A., 2002. Something from "nothing" – eight weak estrogenic chemicals combined at concentrations below NOECs produce significant mixture effects. *Environ. Sci. Technol.* 36, 1751–1756.
- Simmons, S.O., Fan, C.Y., Ramabhadran, R., 2009. Cellular stress response pathway system as a sentinel ensemble in toxicological screening. *Toxicol. Sci.* 111, 202–225.
- Simon, E., van Velzen, M., Brandsma, S.H., Lie, E., Loken, K., de Boer, J., Bytingsvik, J., Jenssen, B.M., Aars, J., Hamers, T., Lamoree, M.H., 2013. Effect-directed analysis to explore the polar bear exposome: identification of thyroid hormone disrupting compounds in plasma. *Environ. Sci. Technol.* 47, 8902–8912.
- Smirnova, L., Harris, G., Leist, M., Hartung, T., 2015. Cellular Resilience. *ALTEX* 32, 247–260.
- Smith, M.T., de la Rosa, R., Daniels, S.J., 2015. Using exposomics to assess cumulative risks and promote health. *Environ. Mol. Mutagen.* 56, 715–723.
- Stallman Brown, E., 2012. Measuring individual exposomes. *Emerg. Sci. Environ. Health Decis.* 1–13.
- Sturla, S.J., Boobis, A.R., FitzGerald, R.E., Hoeng, J., Kavlock, R.J., Schirmer, K., Whelan, M., Wilks, M.F., Peitsch, M.C., 2014. Systems toxicology: from basic research to risk assessment. *Chem. Res. Toxicol.* 27, 314–329.
- Tang, J.Y.M., McCarty, S., Glenn, E., Neale, P.A., Warne, M.S., Escher, B.I., 2013. Mixture effects of organic micropollutants present in water: towards the development of effect-based water quality trigger values for baseline toxicity. *Water Res.* 47, 3300–3314.
- Teeguarden, J.G., Tan, C., Edwards, S., Leonard, J.A., Anderson, K.A., Corley, R.A., Harding, A., Kile, M., S. L.M.S., Stone, D., Waters, K.M., Harper, S., Williams, D.E., Tanguay, R.L., 2016. Completing the link between exposure science and toxicology for improved environmental health decision making: the aggregate exposure pathway framework. *Environ. Sci. Technol.* 50, 4579–4586.
- Tice, R.R., Austin, C.P., Kavlock, R.J., Bucher, J.R., 2013. Improving the human hazard characterization of chemicals: a Tox21 update. *Environ. Health Perspect.* 121, 756–765.
- Tollefsen, K.E., Scholz, S., Cronin, M.T., Edwards, S.W., de Knecht, J., Crofton, K., Garcia-Reyero, N., Hartung, T., Worth, A., Patlewicz, G., 2014. Applying adverse outcome pathways (AOPs) to support integrated approaches to testing and assessment (IATA). *Regul. Toxicol. Pharmacol.* 70, 629–640.
- Toniolo, P., Boffetta, P., Shuker, D., Rothman, N., Hulka, B., Pearce, N., 1997. Application of Biomarkers in Cancer Epidemiology. IARC Scientific Publications No. 142 International Agency for Research on Cancer, Lyon, France.
- Tufi, S., Wassenaar, P.N.H., Osorio, V., De Boer, J., Leonards, P.E.G., Lamoree, M.H., 2016. Pesticide mixture toxicity in surface water extracts in snails (*Lymnaea stagnalis*) by an *in vitro* acetylcholinesterase inhibition assay and metabolomics. *Environ. Sci. Technol.* 50, 3937–3944.
- Vidal-Dorsch, D.E., Bay, S.M., Moore, S., Layton, B., Mehinto, A.C., Vulpe, C.D., Brown-Augustine, M., Loguinov, A., Poynton, H., Garcia-Reyero, N., Perkins, E.J., Escalon, L., Denslow, N.D., Cristina, C.-D.R., Doan, T., Shukradas, S., Bruno, J., Brown, L., Van Agglen, G., Jackman, P., Bauer, M., 2016. Ecotoxicogenomics: microarray interlaboratory comparability. *Chemosphere* 144, 193–200.
- Vineis, P., Schulte, P., McMichael, A.J., 2001. Misconceptions about the use of genetic tests in populations. *Lancet* 357, 709–712.
- Vineis, P., van Veldhoven, K., Chadeau-Hyam, M., Athersuch, T.J., 2013. Advancing the application of omics-based biomarkers in environmental epidemiology. *Environ. Mol. Mutagen.* 54, 461–467.
- Vogs, C., Altenburger, R., 2016. Time-dependent effects in algae for chemicals with different adverse outcome pathways: a novel approach. *Environ. Sci. Technol.* 50, 7770–7780.
- Wetmore, B.A., Wambaugh, J.F., Ferguson, S.S., Li, L., Clewell III, H.J., Judson, R.S., Freeman, K., Bao, W., Sochaski, M.A., Chu, T.-M., Black, M.B., Healy, E., Allen, B., Andersen, M.E., Wolfinger, R.D., Thomas, R.S., 2013. Relative impact of incorporating pharmacokinetics on predicting *in vivo* hazard and mode of action from high-throughput *in vitro* toxicity assays. *Toxicol. Sci.* 132, 327–346.
- Whitesides, G.M., 2015. Reinventing chemistry. *Angew. Chem. Int. Ed.* 54, 3196–3209.
- Wild, C.P., 2005. Complementing the genome with an "exposome": the outstanding challenge of environmental exposure measurement in molecular epidemiology. *Cancer Epidemiol. Biomark. Prev.* 14, 1847–1850.
- Wild, C.P., 2012. The exposome: from concept to utility. *Int. J. Epidemiol.* 41, 24–32.
- Worth, A., Barroso, J., Bremer, S., Burton, J., Casati, S., Coecke, S., Corvi, R., Desprez, B., Dumont, C., Gouliarmou, V., Goumenou, M., Gräpel, R., Griesinger, C., Halder, M., Roi, A.J., Kienzler, A., Madia, F., Munn, S., Nepelska, M., Paini, A., Price, A., Prieto, P., Rolaki, A., Schäffer, M., Triebe, J., Whelan, M., Wittwehr, C., Zuang, V., 2014. Alternative Methods For Regulatory Toxicology – A State-Of-The-Art Review (Available at echa.europa.eu).
- Yugi, K., Kubota, H., Hatano, A., Kuroda, S., 2016. Trans-omics: how to reconstruct biochemical networks across multiple 'omic' layers. *Trends Biotechnol.* <http://dx.doi.org/10.1016/j.tibtech.2015.1012.1013>.